

# Expert Opinion

1. Introduction
2. Self-organised electrochemical formation of nanopore and nanotube structures
3. Nanoporous anodic aluminium oxide
4. Nanotubular titania
5. Other nanoporous and nanotubular metal oxides
6. Expert opinion

**informa**  
healthcare

## Self-ordered nanopore and nanotube platforms for drug delivery applications

Dusan Losic<sup>†</sup> & Spomenka Simovic

*University of South Australia, Ian Wark Research Institute, Mawson Lakes Campus, Mawson Lakes, Adelaide, SA 5095, Australia*

The application of nanotechnology to medicine termed as 'nanomedicine' is recognised as an emerging field with enormous potential for developing new therapeutic concepts. A range of nanoscale materials have been explored in the last few years for drug delivery to address the problems associated with conventional drug therapies such as limited drug solubility, poor biodistribution, lack of selectivity and unfavourable pharmacokinetics. Among them, nanoporous materials with ordered and controlled pore structures, high surface area and pore volume, attracted great attention, particularly for implantable drug delivery systems. This review presents the recent progress in this field focused on electrochemically engineered nanopores/nanotube materials such as nanoporous alumina and nanotubular titania. The basic concept of fabrication of these unique materials using a self-ordering process, description of their structural properties, biocompatibility and recent applications for therapeutic implants is presented.

**Keywords:** drug delivery, electrochemical anodisation, nanoporous alumina, nanotube titania

*Expert Opin. Drug Deliv.* (2009) 6(12):1363-1381

### 1. Introduction

To avoid problems associated with conventional drug therapies related to limited drug solubility, poor biodistribution, lack of selectivity and unfavourable pharmacokinetics, considerable research has been directed in the last two decades towards the development of new and more efficient drug delivery systems [1-3]. Hydrophobic drugs do not dissolve in the blood and do not reach their target, which reduces their pharmacological efficiency [3]. Bioactive agents such as proteins, nucleic acids, enzymes and genes administered through oral or intravenous routes can be degraded prematurely by metabolism or by enzymatic conditions existing in gastrointestinal tracts [3,4]. These challenges have contributed to the development of new controlled drug delivery systems to achieve various goals, such as delivery of therapeutic agents to the desired site, enhancing bioavailability and drug protection.

The application of nanotechnology to medicine, referred to as 'nanomedicine', is recognised as an emerging field with enormous potential for the development of new therapeutic concepts [5-7]. Knowledge of materials at the nanoscale may accelerate the improvement of drug delivery systems, especially in treating life-threatening conditions such as cancer and heart disease [8,9]. A range of new nanoscale materials have been explored in recent years for drug delivery applications, including nanoparticles, nanofibres, dendrimers, liposomes, polymer micelles, carbon nanotubes, fullerenes, nanogels, nanocrystals, viral vectors and virus-like particles (VLP) [5,7-11]. Among them nanoporous and nanotube carriers, owing to their unique features such as low cost fabrication, controllable pore/nanotube structure, tailored surface chemistry, high surface area, high loading capability, chemical resistivity and mechanical rigidity, have engaged a special niche in drug delivery technology [12-14]. Drug

delivery systems based on mesoporous silica prepared by organic synthesis and porous silicon fabricated by electrochemical process have been widely explored in the last few years [12-17]. However, owing to their exceptional properties, increased research interest has recently been focused on electrochemical synthesis of self-organised nanopores and nanotube materials from transition and valve metal oxides [11,14,18-20]. Nanoporous anodic aluminium oxide (AAO) and nanotubular titania ( $\text{TiO}_2$ ) are popular examples, and typical images showing their highly ordered pore and nanotube structures are presented in Figure 1.

This review presents recent developments on applications of self-ordered nanoporous and nanotubular platforms for the delivery of therapeutics, focused primarily on nanoporous alumina and nanotubular titania. First, the phenomenon of the electrochemical formation of nanopores and nanotubes by a self-ordering process is explained. Then fabrication, properties and biocompatibility of these two remarkable materials are addressed briefly, including the current stage of knowledge and recent studies on their implantable drug delivery applications. Finally, examples of nanopore and nanotube materials fabricated from other biocompatible metals and their alloys are discussed.

## 2. Self-organised electrochemical formation of nanopore and nanotube structures

Self-assembly, or the spontaneous organisation of small units into large-scale ordered and stable structures, is ubiquitous in nature and spans a wide range of scales, from the molecular, nano, micro and macro to the planetary scale. Self-assembly and self-organised (or self-ordered) processes are recognised as cost-effective, and an elegant route in nanotechnology for the spontaneous generation, ordering and hierarchical organisation of complex and functional nanomaterials at the molecular and nano scale [21,22]. Several self-assembly synthetic approaches based on chemical, electrochemical, sol-gel and hydrothermal methods have been introduced [22,23]. The recent progress in electrochemical approaches for self-organised formation of nanostructures, such as highly ordered nanoporous alumina and nanotubular titania, placed this method as one of the most popular nanofabrication strategies, with more than 1000 publications in the last 5 years [11,18-20,24-27].

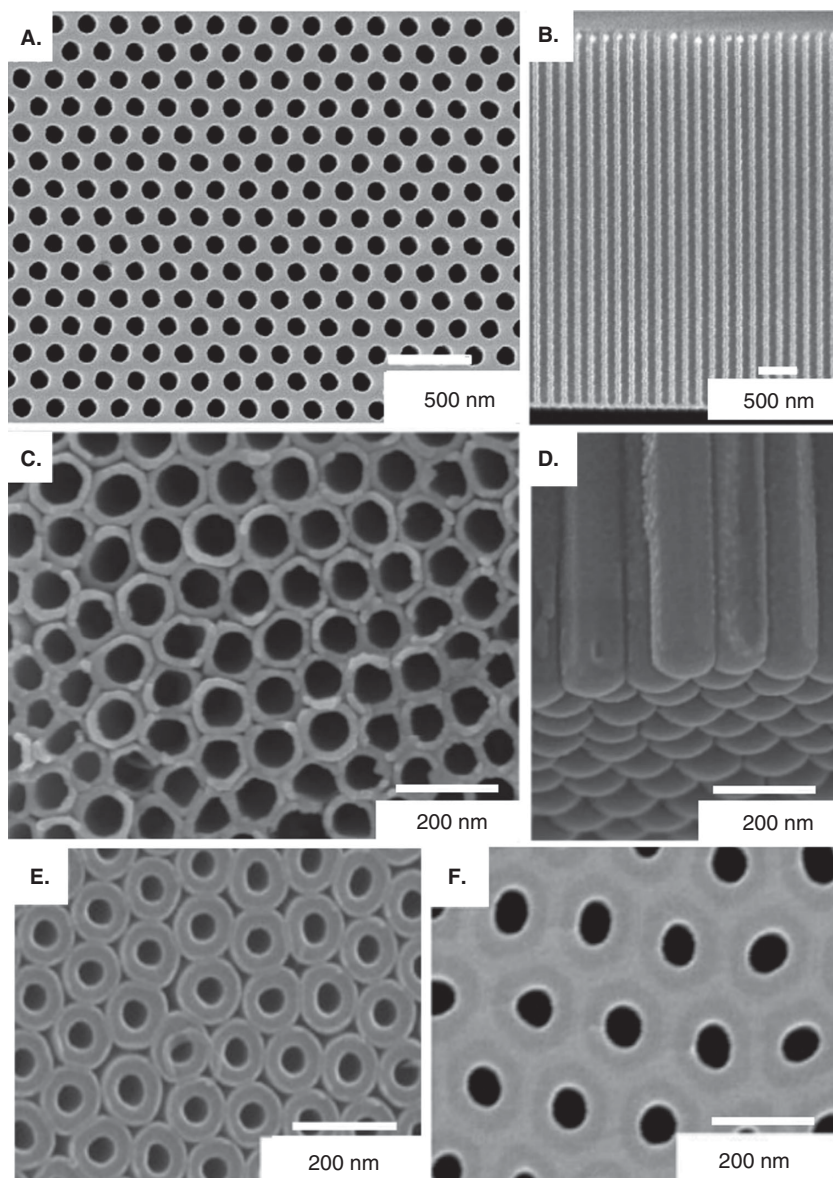
The basic set-up and schematic of the self-ordering process based on electrochemical anodisation is outlined in Figure 2. Anodic oxidation (or anodisation) is an electrochemical process by which an electrochemically active species is oxidised by the passage of current or an applied voltage in an appropriate electrolyte. During this process in an electrochemical cell, the metal serves as the anode connected to the positive terminal of a direct current (DC) power supply, while a chemically stable metal such as platinum and carbon serves as the cathode (Figure 2A). When the circuit is closed, electrons are withdrawn from the metal at the positive terminal, allowing

ions at the metal surface to react with water to form an oxide layer on the metal. The overall electrochemical reactions that occur during electrochemical anodisation of metal are presented in Figure 2A. At the beginning of anodisation, the metal surface is covered with compact and uniform oxide film (Figure 2B, stage I). However, the distribution of the electric field ( $E$ ) in the oxide is strongly influenced by surface morphological fluctuations generating more locally focused electric field (Figure 2B, stage II). As a result of field-enhanced dissolution in oxide films, the pores start to form (Figure 2B, stage III), and finally uniformly distributed pores are formed when the pore growth process reaches a steady-state (Figure 2B, stage IV). The typical current density curve during this process showing these stages is presented in Figure 2C. The final porous oxide layer is formed as a result of the continuous process, which involves competition between dissolution of oxide at the oxide/electrolyte interface and oxidation of metal at the oxide/metal interface [19]. However, these pores are not ordered at the beginning, but during the anodisation process as the pores grow into the bulk metal and improve their arrangement after a long anodisation time. The driving force for self-assembly has been attributed to mechanical stress caused by the repulsive forces between neighbouring pores that lead to ideal self-organisation of AAO with perfect hexagonal arrangement of pores (Figure 1A, B) [26]. Depending on the type of metal, electrolyte and voltage, two typical oxide growth morphologies can be obtained by the self-organising anodisation process, including porous and tubular structures (Figure 3) [19]. Metals such as Al, Nb, Ta and Hf were found to form pore structures, and several other metals (Ti, Ta, Hf, W, Zr, Nb, TiNb, TiZr, TiAl, TiAlNb, etc.) form tubular structures (Figure 3). The formation mechanism for the oxides with nanotubular morphology is governed essentially by the same self-ordering principle and growth mechanism for porous oxide such as highly ordered AAO. Tuning the pore to nanotube morphology has been demonstrated for some metals (Ti, Zr, Ta, Hf) by changing the anodisation parameters such as temperature, applied voltage, electrolyte, pH viscosity and solvent [19].

## 3. Nanoporous anodic aluminium oxide

### 3.1 Fabrication and properties

Nanoporous AAO fabricated by electrochemical anodisation of aluminium is now one of the most popular nanomaterials because of its simple and low-cost fabrication, and remarkable properties such as chemical, thermal stability, hardness, high surface and highly ordered pore structures [18,26-28]. Its applications are diverse and include molecular separation, template synthesis, catalysis, solar cell, energy storage, biosensing, cell growth and drug delivery [26-35]. The structure of AAO can be described as a close-packed hexagonal and perpendicular orientated array of columnar cells, each containing a central pore, of which the size and interval can be controlled by changing the anodisation conditions (Figure 4A) [26,28].



**Figure 1. SEM images of electrochemically fabricated nanoporous and nanotube layers. A, B.** Pore structures of anodic aluminium oxide, top view and cross-section. **C, D.** Nanotube structures of titania, top view and bottom view. **E, F.** Transition from nanopores to nanotubes, self-ordered anodic oxide structures on TiNb, top view.

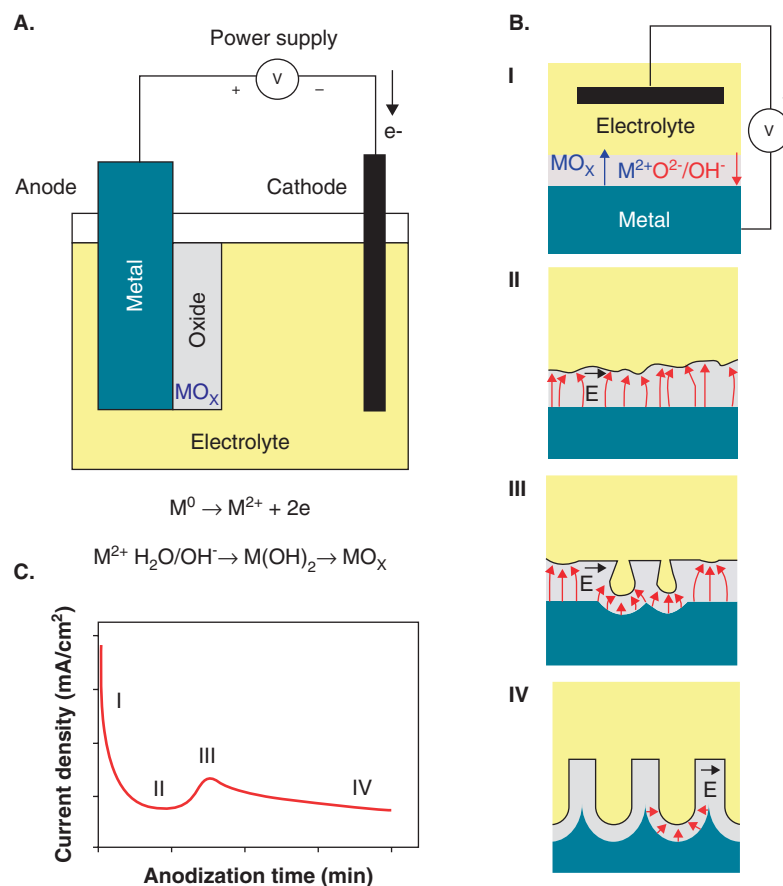
**A, B.** Reproduced with permission from [43].

**C – F.** Reproduced with permission from [19].

Important features are: fully controllable structural dimensions including pore diameter from 10 to 400 nm; interpore distances from 50 to 600 nm; pore aspect ratio from 10 to 5000; the thickness of porous layer from half a micrometre to several hundred micrometres; pore density  $10^9 - 10^{11} \text{ cm}^{-2}$ ; and porosity (5 – 50%) [18,26,35-37]. Fabrication of porous alumina can be performed on conformal to arbitrary surfaces, either flat or curved surfaces, including bulk Al, foils, wires, tubes, and Al films on Si wafers, glass, titanium; and also on various forms such as porous layer on surface and self-supporting membranes. The fabrication process is simple,

low cost, does not need a clean room, and is compatible with existing lithographic and micro-electro-mechanical systems (MEMS) and technologies.

Numerous studies over several decades have explored the anodisation conditions of aluminium, such as voltage, current, electrolyte composition, concentration, temperature and pre-patterning of surface in order to achieve a self-ordering regime and highly ordered AAO pore structures with desired pore diameters [38-41]. In general, three well-studied growth regimes using conventional, so-called mild (MA) or low-field anodisation in  $\text{H}_2\text{SO}_4$  at 25 V,  $\text{H}_2\text{C}_2\text{O}_4$  at 40 V, and  $\text{H}_3\text{PO}_4$



**Figure 2. A schematic diagram showing pore formation by electrochemical self-ordering. A.** Scheme of electrochemical cell for anodisation and corresponding electrochemical reactions. **B.** Scheme of pore formation, which includes several steps: (I) the formation of oxide layer on metal surface; (II) local field distributions caused by surface fluctuations; (III) the initiation of pore growth by field-enhanced dissolution; and (IV) the pore growth in steady-state condition. **C.** Typical current density curve obtained with anodisation showing these stages [19].

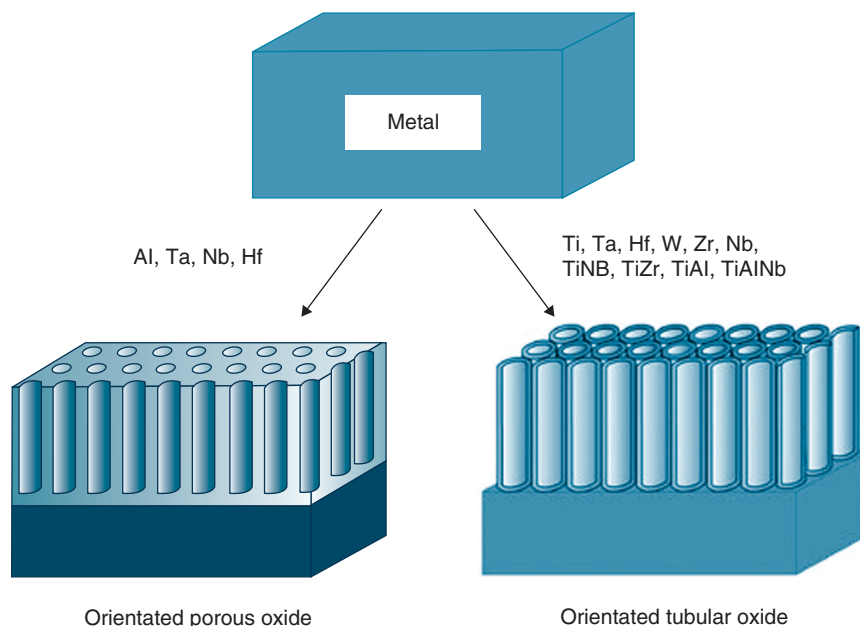
at 194 V, giving 63, 100 and 500 nm interpore distances are accepted as the optimal conditions for fabrication of AAO (Figure 4B). It is generally accepted that the pore diameters ( $D_p$ ) and interpore distance ( $D_c$ ) are linearly proportional to the forming potential of the steady-state growth of anodic porous alumina with a proportionality constant of  $\sim 1.29$  nm/V ( $D_p$ ) and 2.5 nm/V ( $D_c$ ) [26]. The significant improvement of self-ordering process discovered by Masuda and Fakuda in 1995 relies on self-ordering of pores at the bottom of AAO channels after the first anodisation step [42]. This two-step anodisation approach has been a particularly important advance towards simple fabrication of highly ordered porous alumina [42]. By combining the self-ordering process and pre-patterned surface of aluminium using various imprinting techniques to guide pore initiation, it is possible to obtain not only long ordered arrangements but also change from a hexagonal to a square arrangement [43]. One of the disadvantages of mild anodisation is the low rate, typically (1 – 2  $\mu\text{m/h}$ ), which requires a fabrication time of several days. This problem has recently been solved by using a higher current

anodisation condition termed hard anodisation (HA). An anodisation rate of 50 – 100  $\mu\text{m/h}$  has been achieved with a considerably increased speed of fabrication [44,45].

### 3.2 Structural and surface modifications of nanoporous AAO

Several new fabrication processes of nanoporous AAO with tailored pore morphologies and different internal pore geometries have been demonstrated for the preparation of branched, dendritic, hierarchical, multilayered and modulated pore structures [46–48]. An anodisation method called cyclic anodisation has been introduced recently by Losic *et al.* for fabrication of pore structures with different pore geometries and complex pore architectures (Figure 4C) [49,50]. The nanostructuring of internal pore structures is recognised as a useful strategy for a range of applications, including drug delivery. The nanostructured pores could have a considerable impact on diffusion of drug molecules from the pores and therefore provide sustained drug release. The increase in surface area of porous matrix compared with smooth pores is also useful to





**Figure 3. Schematic of two typical morphologies (nanopore and nanotubes) of oxide layers formed by the self-organised anodisation process, which can be obtained from different metals by controlling anodisation parameters such as temperature, applied voltage, electrolyte, pH viscosity and solvent.**

Adapted with permission from [19].

increase the drug loading capacity. The authors emphasise the advantages of these new fabrication strategies for offering new opportunities for designing new nanoporous architectures that can potentially have future impact on the development of new drug delivery systems.

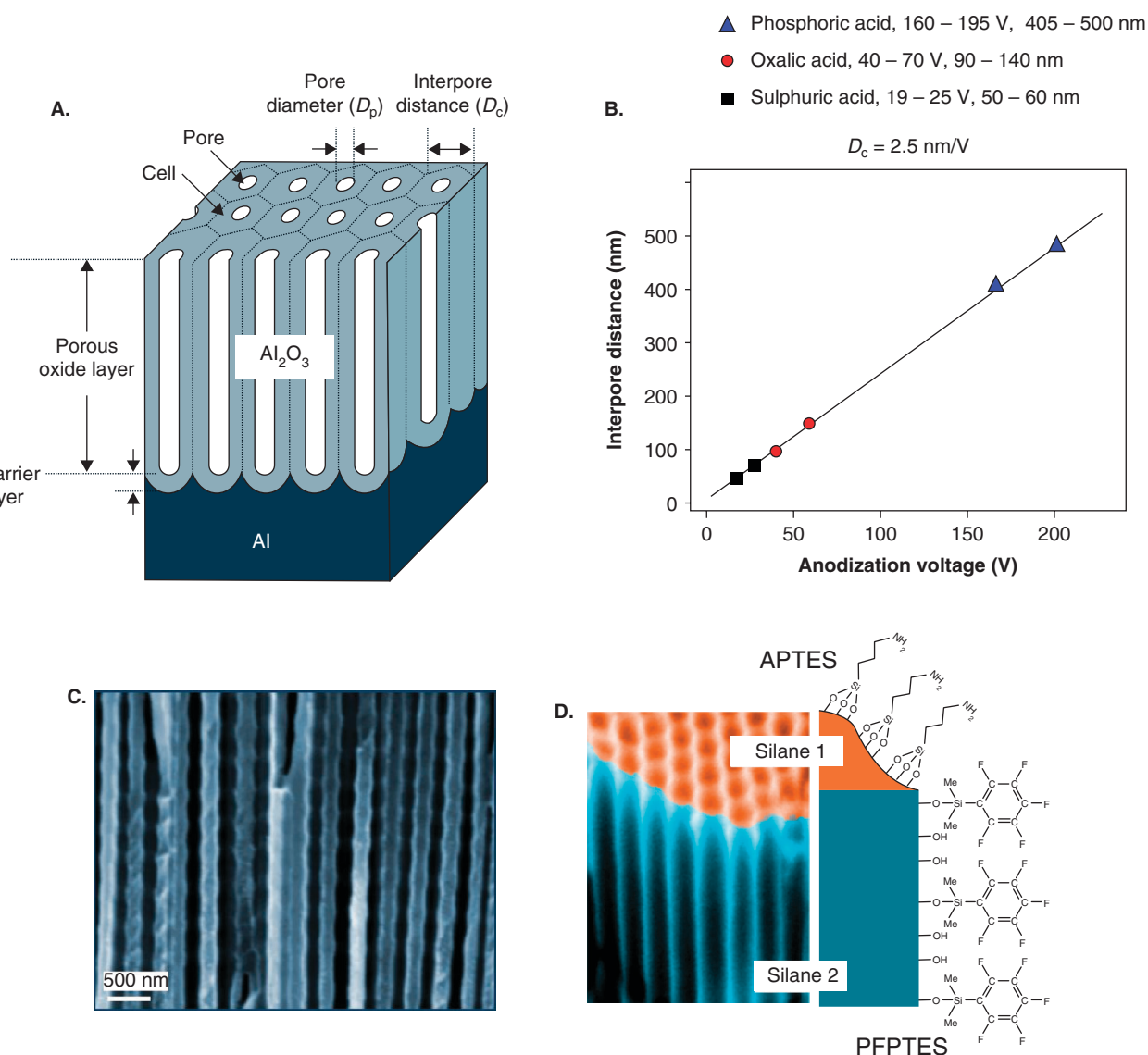
The surface of porous alumina is inherently charged as a consequence of the equilibration of charged crystalline lattice defects on the surface [51]. Depending on the net concentration of lattice defects, the surface may be positively or negatively charged with an attendant redistribution of oppositely charged lattice and electronic defects in the near surface region, which can have a significant impact during interaction with biological materials [51]. Therefore, it is highly desirable to modify the surface of nanoporous AAO and improve biocompatibility for applications that involve interaction with biomolecules and cells, such as protein separation, immunisolation, cell adsorption/growth, tissue engineering and drug delivery. Various solid and soft surface modification techniques of AAO have been explored in the past, including atomic layer deposition (ALD), chemical vapour deposition (CVD), thermal vapour metal deposition, chemical, electrochemical deposition, sol-gel, layer-by-layer deposition and plasma polymerisation [52-55]. A range of surface functionalisations using organic molecules such as silanes, polythiols, *n*-alkanoic acids, polyelectrolyte layers, poly(ethylene glycol) (PEG), chitosan, poly(sodium styrenesulfonate) (PSS), lipid bilayers and plasma polymers have been reported [54-61]. The fabrication of AAO with layered surface chemistry and multifunctional and tailorable properties inside pores is a particularly

important accomplishment towards designing AAO with controlled drug delivery properties (Figure 4C) [62]. Unlike with unmodified pore surfaces, drug molecules can be covalently immobilised in the modified pores or trapped between functional layers. Furthermore, drug release can be controlled by triggering an external stimulus such as pH, UV or temperature. These studies suggest that surface modification can be used as a promising strategy to improve the loading and release properties of AAO with a good prospect of adjusting their properties for specific drug delivery applications [63].

### 3.3 Biocompatibility

Biocompatibility is a prerequisite for the application of new biomaterials and it is defined in terms of cellular response and tissue integration of implantable biomaterials. The bioceramic alumina has been proven as biocompatible and consequently widely used clinically for implants in orthopaedic prostheses and as a dental implant material [64]. The main biocompatibility studies were performed on nanoporous AAO related to their applications for orthopaedic and blood (coronary) implants and immunisolation [65-73].

A major challenge in orthopaedic biomaterials research is to design surfaces that will promote *in vitro* and *in vivo* osteogenesis. However, the influence of surface micro- and nano-topography on osteogenesis is still not well understood. Efforts to use smooth-surfaced alumina have been widely reported in the past, but recent studies have moved towards nanostructured alumina, which shows better bone in-growth [65-70]. The use of nanostructured biomaterials in bone engineering is



**Figure 4.** **A.** The scheme of nanoporous anodic alumina (AAO) structures fabricated by electrochemical anodisation of aluminium. **B.** The dependence of interpore distances and pore diameters of AAO on the electrolyte used (sulphuric, oxalic, phosphoric acid) and anodisation condition (voltage) [44]. **C.** AAO with shaped pore structure fabricated by the cyclic anodisation process, which involves periodic changing of voltage or current during anodisation [49]. **D.** AAO with different surface chemistry on the top and inside of pores [62].

biologically inspired because bone is a naturally occurring porous ceramic material composed of nano-sized organic and mineral phases that form a large macrostructure [74]. Proteins in extracellular bone matrix and calcium phosphate, important constituents of the bone matrix, are nanostructured but the porosities in human bone are predominantly in the range 1 – 100  $\mu\text{m}$ . Earlier studies using oxide ceramics showed that a porous surface with pore diameter of  $\sim 100 \mu\text{m}$  is optimal for bone in-growth to maintain blood supply to connective tissue.

Recent studies using nanoporous alumina, however, have revealed that much smaller pores of AAO allow bone in-growth [65–70]. Study of osteoblast response to nanoporous AAO membranes with pore diameters of 30 – 80 nm confirms

that extending growth of osteoblasts into the nanopores produced an active matrix in the form of fibrous extensions that contain calcium and phosphorus, typical elements of the bone matrix [66]. A similar conclusion was reached by Popat *et al.*, who reported that nanoporous AAO membranes showed significantly improved osteoblast adhesion and proliferation in comparison with amorphous alumina, aluminium and glass [65]. In this case, a nanoporous surface was revealed as a framework in which osteoblasts produce new bone. Further improved osteoblast adhesion and growth on anodised alumina with the same pore diameters (30 – 80 nm) is reported with modified surfaces by physically adsorbing vitronectin or covalently immobilising a cellular adhesive

peptide arginine-glycine-aspartic acid-cysteine (RGDC) [69]. These encouraging studies show the potential of porous AAO for use as a biocompatible platform for bone growth and orthopaedic implant applications, and more studies are expected in the near future.

The interactions between nanoporous AAO with different pore diameters (20 and 200 nm) and whole blood and platelet-rich plasma were investigated by Karlsson *et al.* [70-73]. The blood-implant contact leads to a series of interlinked events such as protein adsorption, complement activation, platelet and leukocyte adhesion/activation, and activation of the coagulation cascade [71-73]. On blood-implant contact, almost instantly the surface is covered with a layer of plasma proteins. Platelet agonists interact with specific receptors on the platelet plasma membrane, generating physiological responses, including: release of intracellular granule contents, change in shape that promotes adhesion, increased affinity to soluble fibrinogen, rearrangements of the membrane phospholipids into procoagulant surface and platelet microparticle (0.1 – 1  $\mu\text{m}$ ) release, often used as markers of platelet activation [73]. Immunocytochemical staining and scanning electron microscopy (SEM) analysis indicate different behaviour of AAO with different pore size in contact with whole blood. AAO membranes with 200 nm pore diameters caused lower complement activation, high platelets and nonsignificant microparticle adsorption. On the contrary, AAO membranes with 20 nm pore diameters caused higher complement activation, negligible platelets and significant microparticle adsorption [72]. It is postulated that different protein adsorption and patterning is created on the surface of AAO depending on the pore size, affecting availability of receptors and binding sites, and also platelet activation. The neutrophilic reaction was shown to depend on the type of protein adsorbed on the surface of AAO membrane (pore size 20 and 200 nm) [71]. Neutrophil activation was seen to a higher extent on the uncoated and fibrinogen-coated AAO surfaces in comparison with serum and collagen-coated alumina surfaces. On the contrary, osteoblast development after 24 h was better on collagen-coated AAO in comparison with uncoated, fibrinogen and serum-coated AAO surfaces.

Earlier, *in vivo* biocompatibility investigations of coronary stents with porous AAO layer using a rabbit restenosis model proved biocompatibility and absence of inflammation [75]. However, recent reports on *in vivo* biocompatibility of therapeutic alumina stent coatings investigated using a pig restenosis model indicated inflammatory response [76,77]. The studies on immunoisolation using capsules with AAO membranes with 75 nm pore diameter showed that AAO is non-toxic and without significant complement activation *in vitro* [67]. However, *in vivo* implantation of these capsules into the peritoneal cavity of rats induces a transient inflammatory response, and PEG is useful in minimising the host response to the materials. Blood vessels are seen in the tissue treated with PEG-modified capsules, which suggests that the major amount of inflammation is due to the procedure itself,

whereas surrounding tissues are relatively undisturbed. Considering that inflammatory response of porous alumina can be minimised by PEG surface functionalisation, further investigations are required to establish whether surface modifications can eliminate or minimise observed inflammatory/restenosis effect.

### 3.4 Implantable drug delivery applications of AAO

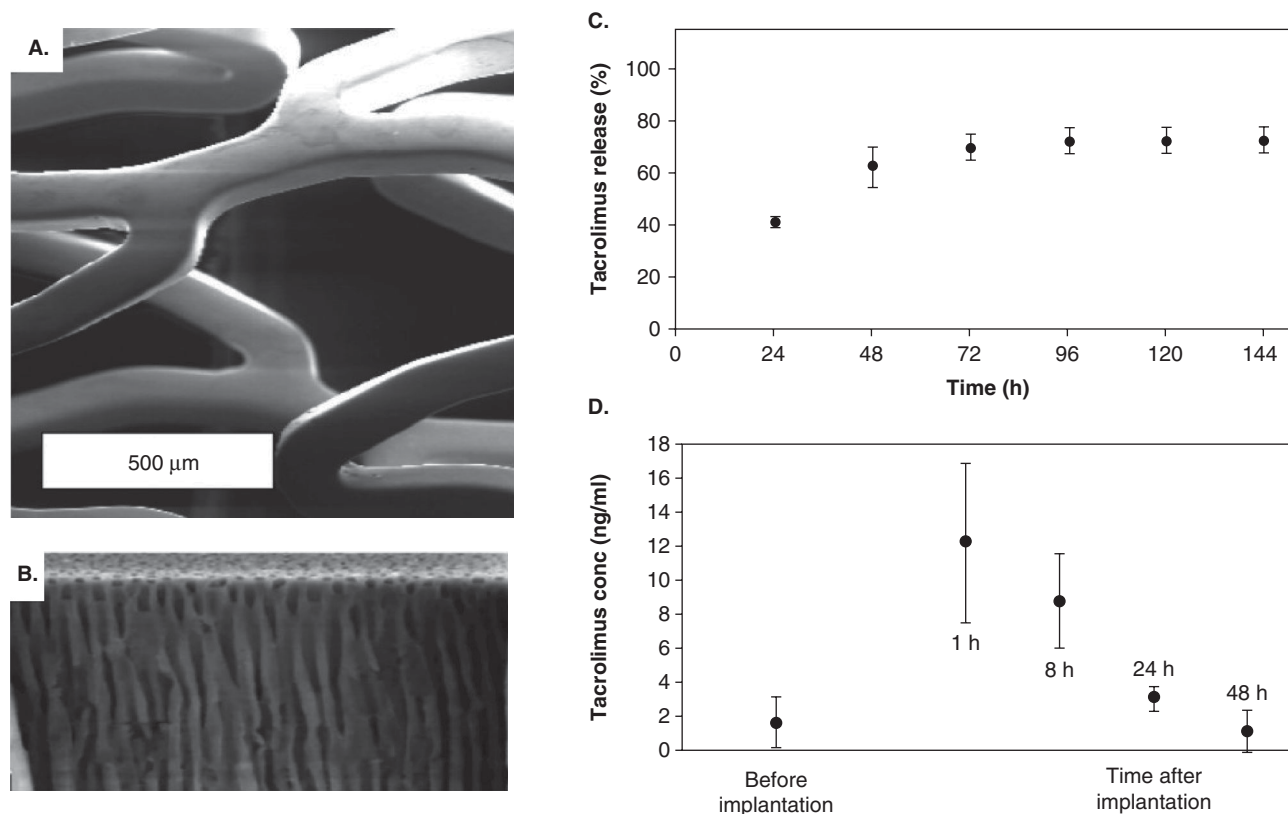
The use of porous alumina for biomedical delivery applications has been explored in several domains including: therapeutic devices for bone and dental tissue engineering, coronary stent implants and carriers for transplanted cells, that is, immunoisolation.

#### 3.4.1 Coronary stents implants

The coronary stent implantation has been established as superior to conventional angioplasty in the treatment of coronary artery disease, but restenosis still remains one of the crucial challenges in interventional cardiology [78,79]. To overcome this major disadvantage, the concept of drug-eluting stents has recently been introduced, where drugs are trapped in polymer films coated on stents, which allows a controlled drug release [78]. Active coating of stents is known to suppress neointima proliferation by releasing anti-inflammatory or antiproliferative therapeutics, for example, immunosuppressive drugs (tacrolimus, sirolimus) and cytostatic drugs (paclitaxel) [78,79]. Although the clinical use of polymer stents has been approved, their inflammatory reaction is still a serious limitation. An alternative approach has been reported recently to design stents with a nanoporous alumina layer filled with anti-inflammatory drugs [75]. The design of active stent coating is one of the most important therapeutic examples for drug release from nanoporous AAO. A SEM image of a stainless-steel stent with a deposited porous alumina layer fabricated by electrochemical anodisation on aluminium film deposited on stainless-steel stent is shown in Figure 5A, B [75]. Wieneke *et al.* investigated alumina nanoporous stent coatings loaded with tacrolimus, an immunosuppressive drug [80]. The effect of AAO pore diameter and depth on the release of 2-deoxyadenosine showing sustained release for up to 40 h with pore diameters ~ 20 nm and further delay was achieved by changing the depth from 1 to 4.4  $\mu\text{m}$  (Figure 5C, D). These results show that stents with an AAO layer could offer alternative solutions by replacing polymer-coated stents and overcome existing problems of inflammatory response and increased neointima formation. The performance of porous AAO stents can be improved further by designing optimal pore structure and surface modification to increase their compatibility and loading capacity, including controlled release.

#### 3.4.2 Biocapsules for immunoisolation

The encapsulation of living cells, that is, cellular immunoisolation using microfabricated capsules with semipermeable barriers, has been investigated over the past few decades as a



**Figure 5.** **A, B.** SEM image of stents coated with nanoporous alumina oxide (cross-section of AAO layer on the top) with pore size between 5 and 15 nm and pore density  $10^{12} \text{ cm}^{-2}$  allowing sufficient drug loading with tacrolimus. **C.** *In vitro* drug release. Cumulative tacrolimus release within the first 144 h. After 72 h, ~ 75% of the loaded 60 μg tacrolimus has been eluted. About 25% is still trapped in the nanoporous AAO coating. **D.** *In vivo* drug release. Time course of whole blood tacrolimus concentration after stent implantation in the carotid arteries of rabbits.

**A, B.** Adapted with permission from [75].

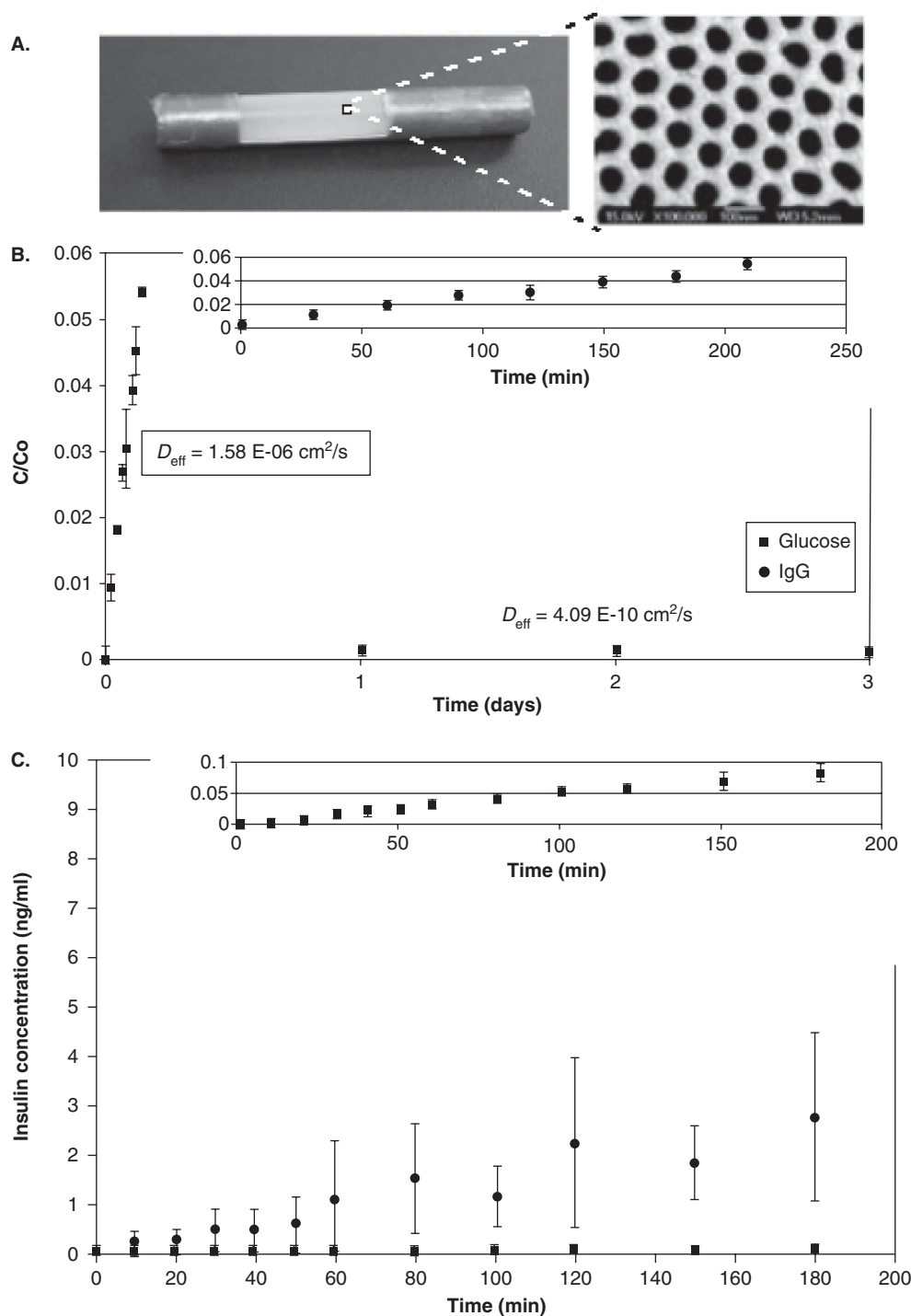
**D.** Adapted with permission from [81].

potential treatment of several diseases, such as diabetes mellitus, Parkinson's and Alzheimer's [67,81,82]. The encapsulation of living cells is a promising future therapy because it has been proved that immunoprotected cells such as pancreatic islets or hepatocytes can respond physiologically both *in vitro* and *in vivo* to appropriate stimuli [83]. Therapeutic cells are encapsulated within a semipermeable membrane that provides the exchange of insulin, oxygen, nutrients and cellular waste; however, larger entities such as immunoglobulins are prevented from penetrating the membrane and destroying the cells. Microfabrication techniques have been applied to create a biocapsule for effective immunoisolation of transplanted cells for treatment of diabetes [81-86]. Typical immunoisolation devices consist of a polymeric membrane, but there are several issues associated with these designs, including poor chemical resistance, inadequate mechanical strength and broad pore size distributions [82,83]. Nanoporous AAO membranes offer many advantages in comparison with polymer membranes in designing implantable biocapsules, such as better mechanical and chemical stability, adjustable

pore size and surface chemistry, and easy incorporation into microfabricated silicon devices.

The use of therapeutic nanoporous AAO biocapsules with controlled drug release for immunoisolation was first demonstrated by Gong *et al.* [86]. Figure 6A shows a capsule fabricated by electrochemical anodisation, consisting of a porous layer with uniform pores with diameters of 25 – 55 nm. To prove the controlled release capacity of these capsules, initial study using model molecules such as fluorescein (molecular mass 400 Da), fluorescein isothiocyanate and dextran conjugates of varying molecular mass as a function of time (molecular mass 4, 20, 70 and 150 kDa) has been performed. The study demonstrates that it is possible to control the diffusion of nutrients and low-molecular-mass proteins (~ 3.5 nm diameter) unhindered, while transport of larger molecules (> 30 nm diameter) was prevented. Their later work demonstrates the feasibility of using nanoporous AAO capsules for the encapsulation of cells. AAO capsules (pore diameter 46 – 75 nm) incorporated with insulin-secreting MIN6 cells could act as effective semipermeable devices,





**Figure 6.** **A.** Photos of completed nanoporous alumina capsule. **B.** Normalised release of glucose (squares,  $D_{\text{eff}} = 1.58 \times 10^{-6} \text{ cm}^2/\text{s}$ ) and IgG (diamonds,  $D_{\text{eff}} = 4.09 \times 10^{-10} \text{ cm}^2/\text{s}$ ) through a nanoporous alumina membrane with a nominal pore size of 75 nm.  $C$  is the concentration at time  $t$ ,  $C_0$  is the loading concentration. Inset: glucose release on a 210-min timescale. **C.** Insulin release from encapsulated (experimental, squares) and unencapsulated (control, diamonds) MIN6 cells following a step increase in glucose. The nominal pore size used for the encapsulated cells was 75 nm. Inset: enlarged graph of experimental data.

Reproduced with permission from [86].

allowing transport of glucose and insulin while impeding the transport of larger proteins such as immunoglobulin G [84-86]. Figure 6B shows fast release of glucose and hindered release of IgG over 4 days. The release profile of insulin through porous alumina from encapsulated MIN6 cells in response to an increase in glucose level is shown in Figure 6C. The experimental trials suggest not only that encapsulated cells do release preformed insulin in response to glucose, but that they are also able to produce new insulin. Furthermore, it is important to emphasise that cells retain their viability in the close proximity of AAO membrane, showing future promise of this approach for clinical immunoisolation.

## 4. Nanotubular titania

### 4.1 Fabrication and properties

Titania nanotube arrays fabricated by a self-ordering process with electrochemical anodisation have also attracted remarkable attention in recent years owing to their unique combination of wide band gap semiconductor properties, nanotube geometry, biocompatibility and large surface area. This material has been used for diverse applications, including photocatalysis for hydrogen production, solar cells, energy storage, catalysis, water purification, sensors, membranes, tissue engineering, implants and drug delivery [19-20,24-25]. Titania nanotube structures, shown schematically in Figure 7A, B, are composed of vertically orientated, highly ordered nanotubes with hexagonal arrangement and controllable nanotube diameters (10 – 300 nm) and thickness (0.5 – 300  $\mu\text{m}$ ). The lengths of the nanotubes correlate with the efficiency of film formation, with the longest nanotubes and the highest efficiency being found for nanotubes formed under controlled voltage. The reason for separation into tubes, as opposed to a nanoporous structure, is not yet entirely clear. A possible explanation is that owing to the electric field and local-heating-enhanced dehydration, the titania nanotubes could separate from each other, where the directions of volume contraction of the hydroxide layer are normal to the walls. Titania nanotubes can be prepared in several forms, including nanotube layer on bulk titanium, self-supporting nanotube arrays and nanotube membranes, and their typical structures are summarised in Figure 7D – G. It is apparent from the cross-sectional SEM images in the inset of the figure that the nanotubes are well separated into individual entities, with an average tube diameter at the top of the layer in the range 160 – 200 nm.

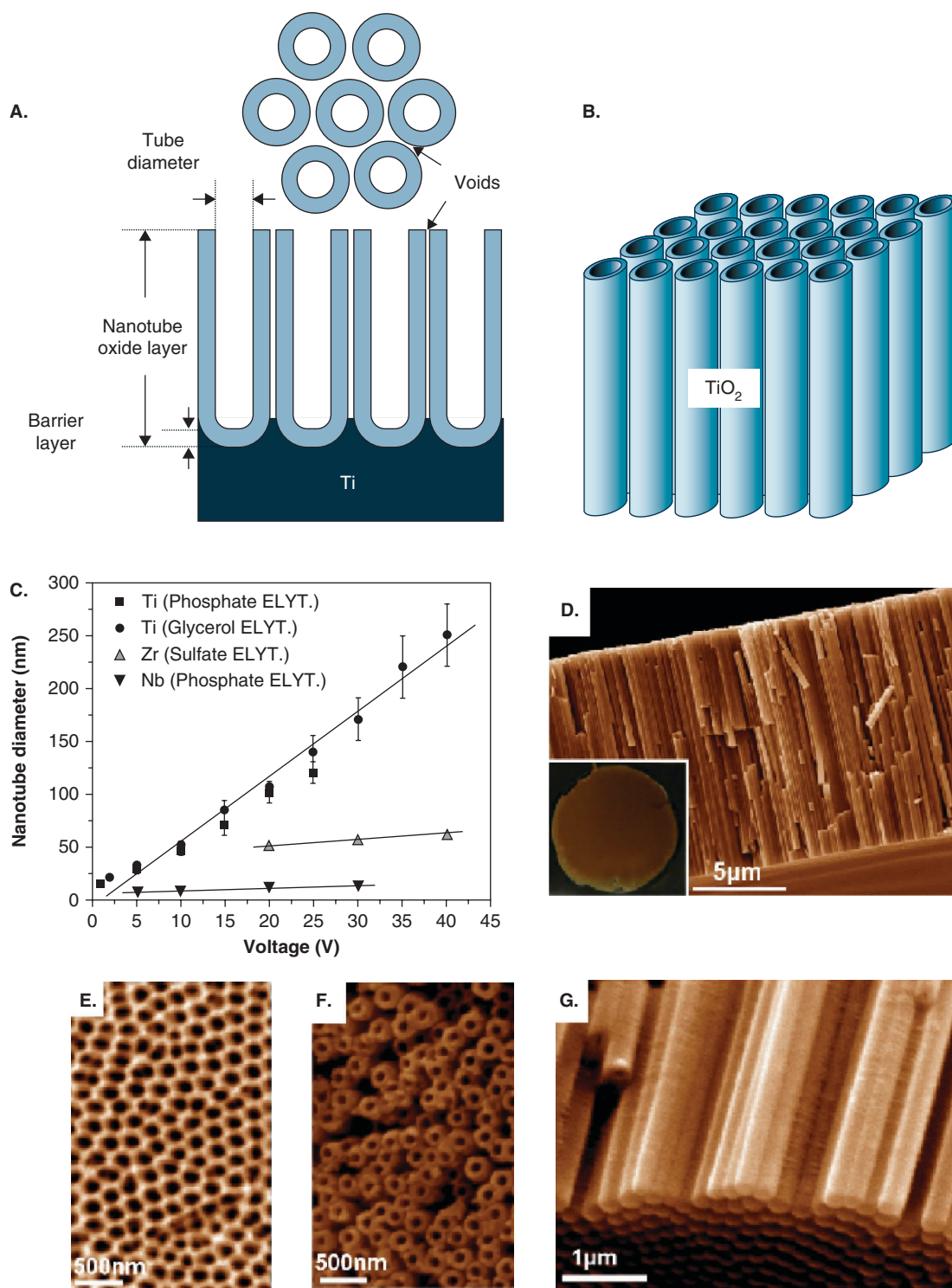
The electrochemical fabrication of self-ordered titania nanostructures was introduced in 1999 by Zwilling *et al.* by anodisation of titanium in a fluoride electrolyte [87]. Since then, several anodisation approaches, mainly focused on finding the optimal electrolyte and anodisation conditions, have been explored to achieve a self-ordering regime for titania nanotube growth [19-20,24-25,88-93]. These studies demonstrated that structural parameters of  $\text{TiO}_2$  nanotubes, including inner diameter, wall thickness, length and  $\text{TiO}_2$  crystallinity, can be

controlled by adjusting electrochemical conditions such as composition of Ti substrate, electrolyte, pH, temperature, anodisation voltage, current and anodisation time (Figure 7C). In general, anodic  $\text{TiO}_2$  nanotube layers can be formed in aqueous and non-aqueous electrolytes containing small amounts of fluoride ions (HF or HF mixtures, NaF or  $\text{NH}_4\text{F}$ ). When anodisation is carried out in acidic, neutral, or alkaline aqueous electrolytes, short tube lengths were formed (500 nm to 2  $\mu\text{m}$ ), which are described as the first generation of titania nanotubes [20,24]. The next generation of nanotubes were grown with thicknesses of > 250  $\mu\text{m}$  in non-aqueous (such as glycerol-based) fluoride-containing electrolytes [19-20,24]. To keep dissolution low and grow long tubes, low electrolyte acidity and low fluoride concentration are desired. The highest tube lengths were obtained in organic viscous electrolytes. This condition under anodisation voltage of 80 – 120 V yields vertically orientated, hexagonally close-packed  $\text{TiO}_2$  nanotube layers, as shown in Figure 1C, D and Figure 7. New self-ordering titania morphologies, such as bamboo-type nanotubes, nanolace, branched tubes, inner tubes and multilayer nanotubes were shown by altering the voltage during the formation of nanotubes [94,95]. Although there is considerable research on surface modification of sister materials such as porous silicon and AAO, surprisingly, the surface modifications of  $\text{TiO}_2$  nanotubes have not yet been widely explored [96].

### 4.2 Biocompatibility

As biocompatible materials and because of their excellent mechanical properties and chemical resistivity, titanium and its alloys, particularly Ti-6Al-4V, have been used extensively in orthopaedic and dental implants since 1970 [97-99]. Most biocompatibility studies on nanotubular titania were focused towards their potential applications for tissue implant engineering, vascular implants and stem cells, where the importance of nanometric scale topography and surface modifications (wettability) were studied.

Several studies by Desai *et al.* have demonstrated that the nanotubular titania surface is a favourable template for bone cell growth and differentiation, and provides clear evidence that osteoblast activity can be significantly enhanced using controlled nanotopographies [100-102]. These surfaces supported higher cell adhesion, proliferation and viability for up to 7 days of culture when compared with plain titanium surfaces [98,101]. Cells cultured on nanotubular surfaces also demonstrated higher alkaline phosphatase activity (ALP) without causing adverse immune response under *in vivo* conditions. *In vivo* biocompatibility was proved by implanting surfaces subcutaneously in rats and performing histological analysis for 4 weeks; chronic inflammation or fibrosis was absent [100]. Increased chondrocyte adhesion on anodised titanium with nanotube structures compared with unanodised titanium has also been reported [103]. Furthermore, the calcium and phosphorous concentrations were 50% higher on these surfaces, suggesting that matrix deposition was



**Figure 7.** **A.** Typical structures of nanotube titania fabricated by electrochemical anodisation of titanium. **B.**  $\text{TiO}_2$  nanotube structures fabricated by electrochemical anodisation in  $\text{NH}_4\text{F}$ /ethyleneglycol electrolyte showing the cross-sectional image of self-supporting  $\text{TiO}_2$  nanotube layer and the entire structure (nanotube film) not detached from titanium substrate (inset). **C, D.** The top of the nanotube surface showing porous and nanotube morphology obtained after removing of the top porous layer. **E.** The top surface showing the nanopores and nanotube structures. **F.** The bottom surface showing the nanotube structures detached from underlying titanium substrate.

upregulated on nanotubular surfaces. The significant adhesion/propagation of the osteoblast by the topography of the TiO<sub>2</sub> nanotubes, with the filopodia of growing cells actually going into the nanotube pores, producing an interlocked cell structure, has been described by Oh *et al.* [104,105]. The growth rate of osteoblast cells is significantly accelerated, > 300 – 400% higher than with Ti surface [104]. *In vivo* studies using implants with 30 nm nanotube diameters placed in the front skull of a domestic pig confirmed the enhanced osteoblast function and bone development [106]. These studies suggest that the incorporation of such nanoarchitectures on implant surfaces has the potential to facilitate the differentiation of cells and promote long-term osteointegration. Owing to their ability to mimic the dimensions of constituent components of natural bone and the possibility to serve as a gene and drug delivery carrier, nanotubes seem to be a promising coating for medical implants.

Two groups recently demonstrated that nanotopography, that is, dimensions of nanotube diameters, of culture media prepared from titania nanotubes can influence adhesion, spreading, growth, and differentiation of human and rat mesenchymal stem cells (MSC) [107-109]. Culturing hMSC on a range of nanotube diameters between 30 and 100 nm, Oh *et al.* found that cell stretching and expression of osteogenic differentiation markers are highest on 100 nm nanotubes [107]. Conversely, Park *et al.* reported different mesenchymal cell response to the nanoscale topography [108]. This study reports the highest adhesion, spreading, growth and differentiation of mesenchymal stem cells on 15 nm nanotubes and dramatically decreased cell functions on 70 and 100 nm nanotubes. Furthermore, Park *et al.* reported similar behaviour of both bone-forming cells and bone-resorbing cells [109]. Considering the same size-dependent response to both bone and mesenchymal cells to TiO<sub>2</sub> nanotubes, it has been suggested that a surface geometry with a lateral spacing of ~ 15 nm that corresponds to the dimension of integrin heads is preferentially recognised by cells. Additionally to nanotube diameters, the surface wettability was also found to be an important parameter for interaction with cells. Bauer *et al.* found considerably enhanced mesenchymal cell attachment to the superhydrophobic titania nanotube surfaces modified by self-assembled monolayers (octadecylphosphonic acid) [110]. The findings of opposite effects of nanotube diameter on osteogenesis of MSCs by two groups motivated more systematic studies on the influence of TiO<sub>2</sub> nanotube geometry, processing parameters, surface chemistry, crystal structure and other differentiation approaches on the response of various stem cells.

As noted previously, vascular prostheses such as stents or vascular grafts are used as common treatments of coronary artery disease. However, such interventions are associated with major complications, such as narrowing of the prosthesis due to vascular smooth muscle cell (VSMC) proliferation and thrombosis as a result of injury and dysfunction of endothelial cells (EC). Active stent coatings with antiproliferative drugs

not only inhibit smooth muscle cells proliferation, but also disturb re-endothelialisation, which increases the risk of thrombosis. TiO<sub>2</sub> nanotubes represent a unique approach where a stent surface can promote re-endothelialisation and decrease VSMC proliferation [102]. When the endothelial layer is denuded, vascular smooth muscle cells may undergo proliferation and intimal hyperplasia, which is the principal mechanism behind restenosis (vessel blockage). The unique effects of titania nanotubes are improved proliferation and function of endothelial cells, decreased proliferation of vascular smooth muscle cells and enhanced production of prostaglandin I<sub>2</sub> (ability to blunt thrombosis and restenosis). The reason for such effects is still unclear, but it is hypothesised that restriction of cell size may play a role. Further studies with different tube dimensions and topographies are warranted in order to understand mechanisms of cell behaviour. These results show that titania nanotubes have various desirable effects on cells involved in repair after vascular injury and are a promising candidate for next generation vascular materials and stent applications.

### 4.3 Drug delivery applications of titania nanotube material

Nanotubular titania for drug delivery applications has been explored mainly for medical implants, such as drug-eluting coating for orthopaedic implants, dental implants and vascular (coronary) stents, where not only the controlled release of drugs, for example, antibiotics or growth factors, is desired, but also appropriate biointegration required [100-106,111]. More recently, stimuli-responsive therapeutic systems based on titania nanotubes have been reported as another promising application.

#### 4.3.1 Therapeutic bone and stent implants

Most medical implant procedures such as hip replacements, dental implants, or vascular stents require subsequent drug therapy to prevent infection, control clotting, or decrease inflammation [112]. Delivery of drugs locally from an implant surface rather than systemically can reduce side effects. A common strategy is drug elution through a drug-loaded polymer coating. Although effective in delivering a drug for long periods, polymer degradation may induce an inflammatory response, activating phagocytes and increasing vascular smooth muscle proliferation, which can lead to implant failure [78]. To address these problems, the newest developments in drug-eluting surfaces are focused on non-polymer-based drug delivery implant platforms, such as nanoporous surfaces (AAO or titania) [102]. These surfaces are capable of eluting drugs and are effective in reducing intimal hyperplasia [113].

Bacterial infection is the most common complication after orthopaedic implant surgery [113]. Bone marrow swells during infection and reduces blood supply because of the compression of blood vessels in the bone marrow. The infection can also spread to the surrounding muscle tissues. To reduce the



chances of serious complications, antibiotic therapy is indicated 6 – 8 weeks after surgery. Systemic antibiotic applications (intravenous, intramuscular, oral) can be toxic and ineffective at reaching infected tissues near the implant. Design of therapeutic implant devices that contain antibiotics (e.g., gentamicin) has several advantages, such as fewer side effects and efficient therapeutic effects. To address the problem of bacterial infection after orthopaedic implant surgery, Popat *et al.* explored titania nanotubular arrays for local delivery of antibiotics off-implant at the site of implantation [102]. A titania nanotube (80 nm pore diameter and 400 nm length) was loaded with gentamicin (70 – 85% loading efficiency). Drug release kinetics are dependent on initial loading; and there was a sustained release, for 45, 90 and 150 min for loadings 200, 400 and 600 µg. Bacterial adhesion on the surface of gentamicin-loaded titania nanotubes is reduced significantly (in comparison with titanium surface and unloaded nanotubes), whereas normal osteoblast adhesion and proliferation is retained [103-111]. Antoci *et al.* came to a similar conclusion, showing that vancomycin covalently tethered to a Ti alloy surface prevents bacterial colonisation and biofilm formation [114,115]. Moreover, the tethered antibiotic is stable, maintains its activity even when challenged multiple times with bacteria and does not foster resistance. Covalent attachment of antibacterial molecules on implant surfaces is advantageous owing to the permanent defence from infection and implant surgery complications.

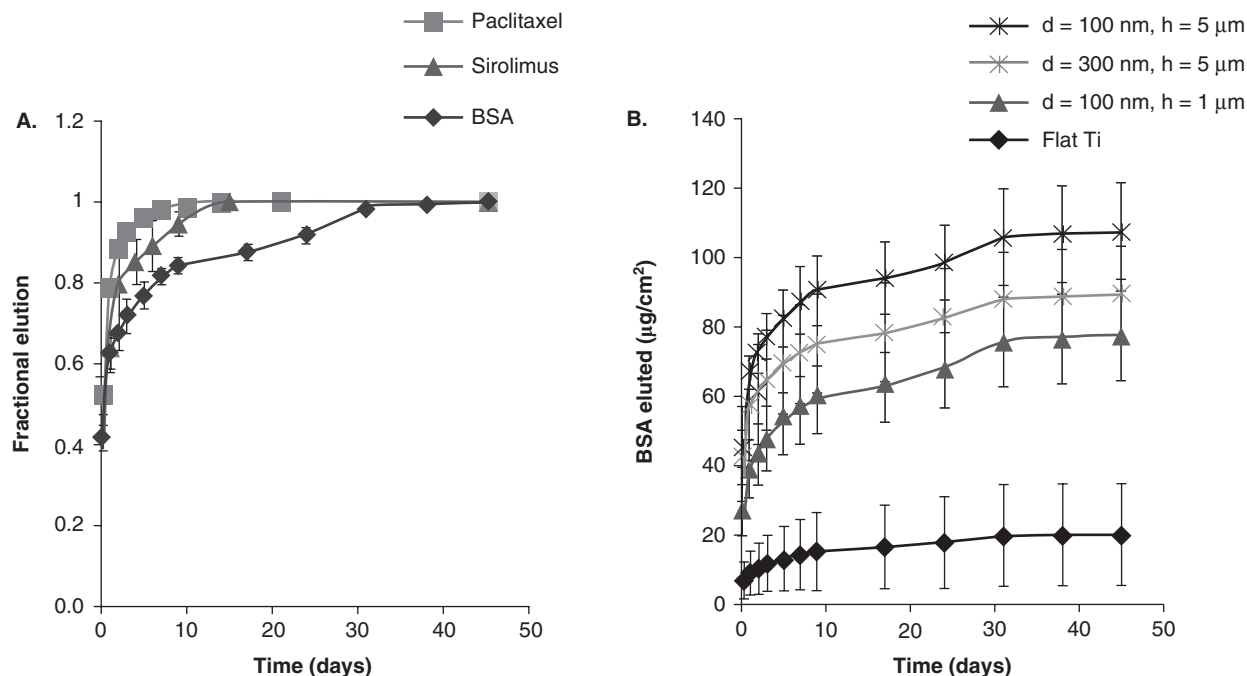
Peng *et al.* recently reported elution kinetics for two model drugs, paclitaxel (11.3 kDa drug with a small (< 0.5 nm) hydrodynamic radius) and bovine serum albumine (66 kDa and hydrodynamic radius 2 – 3 nm) from titania nanotubes with varying length and pore diameter [111]. This study demonstrated that TiO<sub>2</sub> nanotubes can control small molecule delivery in the order of weeks, and larger molecules in the order of months. The total drug elution was mostly affected by nanotube length, and maximum paclitaxel elution was reached at ~ 2 weeks. Comparison between nanotube arrays with the same diameter (100 nm) but different lengths (1 µm versus 5 µm) revealed that on average nanotubes of 1 µm held less than half the amount of drug trapped by 5 µm nanotubes. While nanotubes controlled the elution of small molecules for 7 - 14 days (Figure 8A), they were capable of controlling the delivery of larger molecules such as bovine serum albumin (BSA) for > 30 days (Figure 8B). Data fitted in Fick's first law of diffusion suggest that elution can be described in a two-phase kinetic model, with an initial rapid phase within the first 24 h and a slower phase thereafter. At size scales of 100 nm and larger, diffusion of both types of molecule was largely insensitive to tube diameter, but total drug elution was dependent on tube length, with longer tubes performing better than shorter ones [111].

To demonstrate the efficacy of using titania nanotubes as drug-eluting coatings for implantable devices, Popat *et al.* studied the loading with various amounts of drugs such as BSA (negatively charged) and lysozyme (LYS) (positively charged),

and their controlled release by varying the tube length, diameter and wall thickness [102]. The authors showed that the release rates of BSA and LYS can be controlled by varying the amount of proteins loaded into the nanotubes. Furthermore, by changing the nanotube pore diameter, wall thickness and length, the release kinetics can be altered for each specific drug to achieve a sustained release. The release kinetics was slower and sustained (up to 65 and 110 min) when loading level was increased and electrostatic interaction lysozyme–titania surface operative [102].

#### 4.3.2 Stimuli-responsive therapeutic systems

In conventional porous systems described in the previous sections, the release of adsorbed molecules usually follows a sustained kinetic mechanism that can be expressed in terms of diffusion of adsorbed molecules. It is desirable for certain applications to control release by environmental stimuli such as pH, temperature changes, light or magnetic field [12]. The nanoscale encapsulation of ferromagnetic structures has received a great deal of attention because of the exciting possibilities of using these materials in various applications that range from new electromagnetic to biomedical devices. For example, nanoscale magnetic entities could be transported and concentrated at pre-targeted locations or organs within the human body by means of an external magnetic field in order to exert a specific function with high local and temporal precision. Shrestha *et al.* reported fabrication of magnetically guided titania nanotubes and demonstrated the use of these nanotube layers as magnetically guided photocatalysts important in responsive stimulus release [116]. The drug release concept is based on the fact that the UV-induced hole generation in the valence band of the TiO<sub>2</sub> will lead to chain-scission of a monolayer attached to TiO<sub>2</sub>. As outlined in Figure 9A, the cleavage takes place at the anchoring siloxane groups, which causes the release of the model drug. It should be pointed out that this release approach is almost unique to TiO<sub>2</sub>, as its bond-breaking ability for attached linker molecules occurs because of the position of the TiO<sub>2</sub> valence band relative to the water redox levels. Fluorescence microscopy images demonstrated that UV-induced photocatalytic activity of the magnetic TiO<sub>2</sub> nanotubes may be used to kill cancer cells (Figure 9B, C). Another extraordinary example of using titania nanotubes as a therapeutic agent for the treatment of cancer cells opens many new opportunities for drug delivery applications, which need to be explored in the future [117]. Song *et al.* reported the fabrication and use of an amphiphilic TiO<sub>2</sub> nanotubular structure that provides a highly controllable drug release system based on a hydrophobic cap on a hydrophilic TiO<sub>2</sub> nanotube [96]. This hydrophobic cap prevents uncontrolled leaching of the hydrophilic drug into an aqueous environment. By exploiting the photocatalytic nature of TiO<sub>2</sub> for UV-induced chain-scission of attached organic monolayers, the cap can be removed and a highly controlled release of drugs and proteins can be achieved. The chain-scission-induced release from TiO<sub>2</sub> surfaces



**Figure 8. Small molecules and protein elution study from titania nanotubes.** A. Fractional elution of paclitaxel, sirolimus and BSA. B. BSA elution from titania nanotube arrays of various dimensions. Reproduced with permission from [111].

can be triggered not only by UV light but also by suitable X-ray radiation.

## 5. Other nanoporous and nanotubular metal oxides

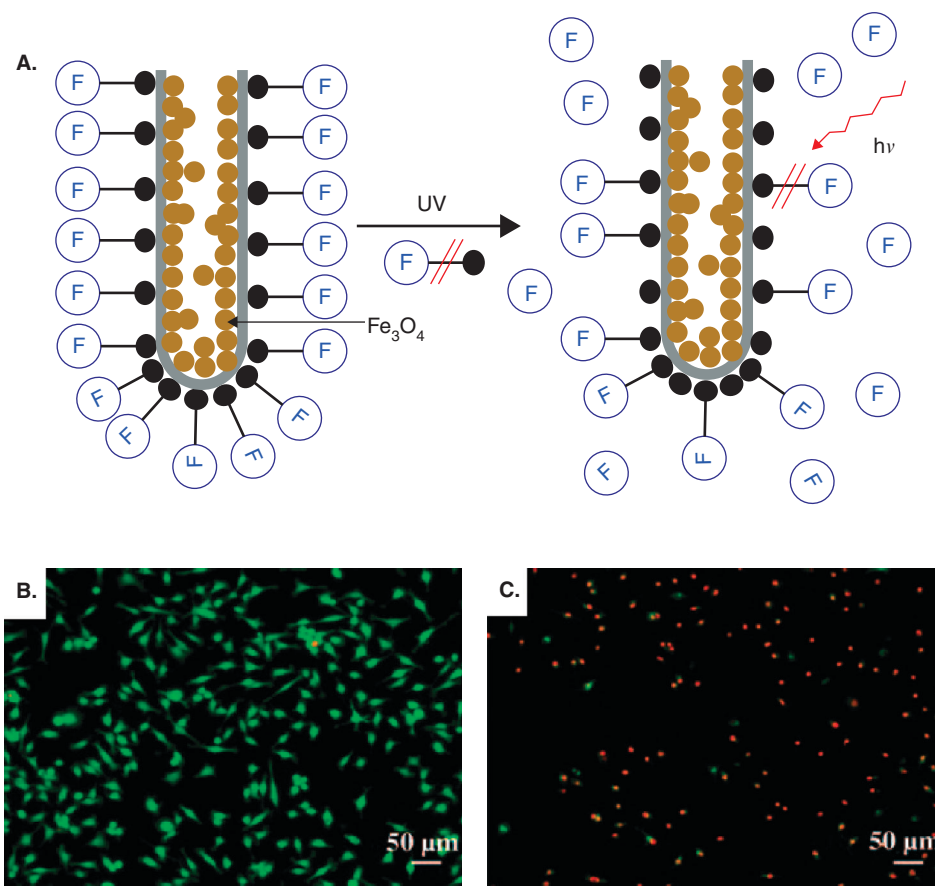
The strategies described previously for self-ordering conditions for growth of oxide with porous and nanotubular morphology have been described for several transitional and valve metals, such as Al, Ti, Ta, W, Nb, Hf, Zr and their alloys TiNb, TiAl, TiZr, Ti6Al7Nb, Ti6Al4V, and so on [19]. The formation of both morphologies is essentially governed by the same self-ordering process. For titanium and another valve metals, in contrast to aluminium, a low pH (acidic electrolyte) is not sufficient to create a porous layer, and the presence of fluoride ions is required to form soluble metal fluoride complex. A chemical dissolution process (in the case of Ti, the HF electrolyte) is suggested to play a key role for separation of nanopores and the formation of nanotubes rather than a nanoporous structure. Extensive research is now underway and more metals can be expected to be fabricated by changing electrolytes and anodisation conditions in order to obtain nanoporous, nanotubular or even mixed morphologies [19].

It has been reported that aluminium and vanadium ions can be dissolved from Ti alloy (Ti-6Al-4V), which is a concern because aluminium is a growth inhibitor of bone and a possible cause of Alzheimer's disease and vanadium has strong cytotoxicity [101,118,119]. Matsuno *et al.* reported the biocompatibility and osteogenesis of several refractory

metal implants (Ti, Hf, Nb, Ta and Re) by both histological examination and imaging methods in *in vivo* animal implantation tests [98]. Titanium, hafnium, niobium, tantalum and rhenium wires were implanted in the subcutaneous tissue of the abdominal region and in femoral bone marrow of rats for either 2 or 4 weeks [119]. No inflammatory response was observed around the implants, and all the implants were encapsulated with thin fibrous connective tissue. No dissolution of these metals was detected by X-ray scanning analytical microscope (XSAM) in the soft tissue [119]. Histological examination of the hard tissue showed that the amount of new bone formation decreased slightly from the second to the fourth week after implantation, and that the percentage of bone in contact with the implant increased markedly over the same period. These results indicate that titanium, hafnium, niobium, tantalum and rhenium have good biocompatibility and osteoconductivity.

The striking possibility for future application of therapeutic titania and other metal oxide nanotubes lies in non-invasive anticancer therapy [116,117]. One of the longstanding problems in medicine is how to cure cancer without harming normal body tissue. The real benefit would be finding a way selectively to kill cancer cells and not damage healthy ones. New devices based on stimuli-responsive titania nanotubes (e.g., magnetically guided, UV, X-ray responsive) represent a promising approach in chemotherapy.

Furthermore, nanopore and nanotube platforms can be prepared in a variety of forms (foils, films and bulk), with capacity to load a large amount of drugs. Titania nanotube



**Figure 9. A.** Diagram showing the release principle of active molecules (model drug) from the functionalised magnetic  $\text{TiO}_2$  nanotubes on irradiation with UV light. A fluorescent dye (active molecule) was attached to the  $\text{TiO}_2$  nanotubes with a siloxane linker. Fluorescence microscopy images demonstrate UV-induced cancer cell killing by using HeLa tumour cells cultured on  $\text{TiO}_2$  nanotube layers. The image shows cells **(B)** without and **(C)** after UV light irradiation. Live cells were detected with calcein and dead cells were detected with EthD-1.

Reproduced with permission from [116].

films, for example, could potentially provide simple and effective alternatives for dermal/transdermal delivery of problematic drugs. Many candidates for dermal/transdermal application (e.g., vitamins, antioxidants) have poor stability against light, oxidation or physiological pH, and consequently poor bioavailability [7]. To overcome this problem, liposomes or emulsions are often applied, which are unstable systems and offer only limited solution, and the use of nanotube platforms could provide many advantages.

## 6. Expert opinion

This review has presented the recent progress on electrochemically engineered nanopore/nanotube materials such as nanoporous alumina and nanotubular titania for implantable drug delivery systems. These new nanomaterials have been shown to have many favourable properties for drug delivery, including high surface area, and controllable pore and nanodube dimensions, geometries and surface chemistry. Owing to their

ability to mimic the dimensions of constituent components of natural bone and the possibility to serve as gene and drug delivery carriers, nanotubes are seen as a promising coating for medical implants. Therefore, it is not surprising that research on applications of porous alumina and nanotubular titania for therapeutic implant devices has been increasing considerably in the past few years. This new field can be described as one of the best examples of multidisciplinary approaches that combine the areas of nanotechnology, biomaterial chemistry, controlled drug delivery and biomedical engineering. Although still in the early stages, a few *in vivo* studies clearly show the potential of these materials for drug delivery devices in orthopaedic implants, dental implants and vascular stents, where not only the controlled release of drugs such as antibiotics or growth factors is desired, but also appropriate biointegration is needed. This is only the beginning of further research in terms of correlating biomaterial chemistry and tissue responses and new clinical approaches required not only for orthopaedics, but also for the treatment of several other

diseases (hearth, cancer, diabetes, Parkinson's, Alzheimer's, etc.). The remarkable catalytic and anticancer therapeutic properties of some of these materials, such as titania nanotubes, are expected to attract more research in the near future.

Future research potentially has multiple directions, including: i) the increasing biocompatibility by covalent immobilisation of therapeutic molecules on implant surfaces, for example, antibacterial and anti-inflammatory drugs; ii) the achievement of controlled and sustained drug elution over a long period of time using specific surface modification, structural design of pores and combining with biocompatible polymers; and iii) the development of stimuli-responsive systems such as magnetic field, light, temperature, pH is particularly important for highly toxic drugs. The implementation of nanoporous and nanotube

membranes or films into microfabrication devices for creating chip-based drug delivery platforms is also a future direction. Therapeutic systems that possess a combination of structural, mechanical, biosensing and electronic features may overcome challenges associated with conventional delivery therapies. Dermal (topical) or transdermal delivery systems are another field of research where these materials could contribute significantly to medical applications.

## Declaration of interest

D Losic and S Simovic are presently involved in a project funded by the Australian Research Council and University of South Australia.

## Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

1. Drews J. Drug discovery: a historical perspective. *Science* 2000;287:1960-4
2. Mainardes RM Silva LP. Drug delivery systems: past, present, and future. *Curr Drug Targets* 2004;5:449-55
3. Fahr A, Liu X. Drug delivery strategies for poorly water-soluble drugs. *Expert Opin Drug Deliv* 2007;4:403-16
4. El-Anead A. An overview of current delivery systems in cancer gene therapy. *J Controlled Release* 2004; 94:1-14
5. Amiji MM. Nanotechnology for targeted drug and gene delivery. *Nanomed Nanotechnol Biol Med* 2006;2:299-300
6. Kayser O, Lemke A, Trejo NH. The impact of nanobiotechnology on the development of new drug delivery systems. *Curr Pharm Biotechnol* 2005;6:3-5
7. De Villiers MM, Aramwit P, Kwon GS. Eds. *Nanotechnology in drug delivery*. Springer, AAPS Press 2009
8. Van D, McGuire T, Langer R. Small scale systems for in vivo drug delivery. *Nat Biotechnol* 2003;21:1184-91
9. Wei C, Wei W, Moris M, et al. Nanomedicine for drug delivery. *Med. Clin N Am* 2007;91:863-70
10. Hughes GA. Nanostructure-mediated drug delivery. *Nanomedicine, Nanotechnology, Biology and Medicine* 2005;1:22-30
11. Martin CR Kohli P. The emerging field of nanotube biotechnology. *Nat Rev Drug Discov* 2003;2:29-37
12. Vallet-Regí M, Balas F, Arcos D. Mesoporous Materials for Drug Delivery. *Angew Chem Int Ed* 2007;46:7548-58
13. Wang S. Ordered mesoporous materials for drug delivery. *Microporous Mesoporous Mater* 2009;117:1-9
- Review on mesoporous materials for drug delivery.
14. Son SJ, Bai X, Nan A, et al. Template synthesis of multifunctional nanotubes for controlled release. *J Controlled Release* 2006;114:143-52
- Review on synthetic strategies for multifunctional nanotubes from porous alumina and their applications for drug delivery.
15. Anglin EJ, Cheng LY, Freeman WR, Sailor MJ. Porous silicon in drug delivery devices and materials. *Adv Drug Delivery Rev* 2008;60:1266-77
- Review on porous silicon for drug delivery.
16. Salonen J, Kaukonen AM, Hirvonen J, Lehto V-P. Mesoporous silicon in drug delivery applications. *J Pharm Sci* 2008;97:632-53
17. Prestige CA, Barnes TJ, Lau C-H, et al. Mesoporous silicon: a platform for the delivery of therapeutics. *Expert Opin Drug Deliv* 2007;4:101-10
- A review on porous silicon for drug delivery.
18. Schmid G. Materials in nanoporous alumina. *J Mater Chem* 2002;12:1231-8
19. Ghicov A, Schmuki P. Self-ordering electrochemistry: a review on growth and functionality of TiO<sub>2</sub> nanotubes and other self-aligned MOx structures. *Chem Commun* 2009;20:2791-808
- Excellent review on self-ordering chemistry approach for synthesis of nanopores and nanotube materials.
20. Grimes CA. Synthesis and application of highly ordered arrays of TiO<sub>2</sub> nanotubes. *J Mater Chem* 2007;17:1451-7
- A review on synthesis and application of titania nanotubes.
21. Colfen H, Mann S. Higher-order organization by mesoscale self-assembly and transformation of hybrid nanostructures. *Angew Chem Int Ed* 2003;42:2350-64
22. Gomar-Nadal E, Puigmartí-Luis J, Amabilino DB. Assembly of functional molecular nanostructures on surface. *Chem Soc Rev* 2008;37:490-504
23. Gooding JJ, Mearns F, Yang W, Liu Jinguang. Self-assembled monolayers into the 21st century: recent advances and applications. *Electroanalysis* 2003;15:83-96
24. Mor GK, Varghese OK, Paulose M, et al. A review on highly ordered, vertically oriented TiO<sub>2</sub> nanotube arrays: Fabrication, material properties, and solar energy applications. *Sol Energy Mater Sol Cells* 2006;90:2011-75
- A review on synthesis and application of titania nanotubes.
25. Macak JM, Tsuchiya H, Ghicov A, et al. TiO<sub>2</sub> nanotubes: Self-organized electrochemical formation, properties and applications. *Curr Opin Solid State Mater Sci* 2007;11:3-18
- A review on synthesis and application of titania nanotubes.



26. Eftekhari A (ed) Nanostructured Materials in electrochemistry. Sulka GD. Highly ordered anodic porous alumina formation by self organized anodizing, Wiley-VCH2008
- **Comprehensive book chapter on anodic porous alumina.**
27. Li L, Koshizaki N, Li G, Nanotube arrays in porous alumina membranes. *J Mater Sci Technol* 2008;24:550-62
28. Thompson GE. Porous anodic alumina: Fabrication, characterization and applications. *Thin Solid Films* 1997;297:192-201
29. Digle JW, Downie TC, Goulding CW, Anodic oxide films on aluminium. *Chem Rev* 1969; 69:365-405
30. Lei Y, Cai WP, Wilde G. Highly ordered nanostructures with tunable size, shape and properties: A new way to surface nano-patterning using ultra-thin alumina masks. *Prog Mater Sci* 2007;52:465-539
31. Takmakov P, Vlasiouk I, Smirnov S. Application of anodized aluminum in fluorescence detection of biological species. *Anal Bioanal Chem* 2006;385:954-8
32. Steinhart M, Wehrspohn RB, Gosele U, Wendorff JH. Nanotubes by template wetting: A modular assembly system. *Angew Chem Int Ed* 2004;43:1334-44
33. Velleman L, Shapter JG, Losic D. Gold nanotube membranes functionalised with fluorinated thiols for selective molecular transport. *J Membr Sci* 2009;328:121-6
34. Losic D, Shapter JG, Mitchell JG, Voelcker NH. Fabrication of gold nanorod arrays by templating from porous alumina. *Nanotechnology* 2005;16:2275-81
35. Lillo M, Losic D. Ion-beam pore opening of porous anodic alumina: The formation of single nanopore and nanopore arrays. *Mater Lett* 2009;63:457-60
36. Ono S, Saito M, Asoh H. Self-ordering of anodic porous alumina formed in organic acid electrolytes. *Electrochim Acta* 2005;51:827-33
37. Jessensky O, Muller F, Gosele U. Self-organized formation of hexagonal pore arrays in anodic alumina. *App Phys Lett* 1998;72:1173-5
38. Choi J, Wehrspohn RB, Gosele U. Mechanism of guided self-organization producing quasi-monodomain porous alumina. *Electrochim Acta* 2005;50:2591-5
39. Masuda H, Hasegawa F, Ono S. Self-ordering of cell arrangement of anodic porous alumina formed in sulfuric acid solution. *J Electrochem Soc* 1997;144:L127-30
40. Schneider JJ, Engstler N, Budna KP, et al. Freestanding, highly flexible, large area, nanoporous alumina membranes with complete through-hole pore morphology. *Eur J Inorg Chem* 2005;12:2352-9
41. Vrublevsky I, Parkoun V, Schreckenbach J. Analysis of porous oxide film growth on aluminum in phosphoric acid using re-anodizing technique. *App Surf Sci* 2005;242:333-8
42. Masuda H, Fukuda K. Ordered metal nanohole arrays made by a 2-step replication of honeycomb structures of anodic alumina. *Science* 1995;268:1466-8
- **The first demonstration of two-step fabrication of porous alumina with highly ordered pore structures.**
43. Masuda H, Asoh H, Watanabe M, et al. Square and triangular nanohole array architectures in anodic alumina. *Adv Mater* 2001;13:189-92
44. Lee W, Ji R, Gosele U, Nielsch K. Fast fabrication of long-range ordered porous alumina membranes by hard anodization. *Nat Mater* 2006;5:741-7
45. Chu SZ, Wada K, Inoue S, et al. Fabrication of ideally ordered nanoporous alumina films and integrated alumina nanotubule arrays by high-field anodization. *Adv Mater* 2005;17:2115-21
46. Meng G, Jung YJ, Cao A, Vajtai R, Ajayan PM. Controlled fabrication of hierarchically branched nanopores, nanotubes and nanowires *PNAS* 2005;102:7074-7078
47. Ho AYY, Gao H, Lam YC, Rodriguez I. Controlled fabrication of multitiered three-dimensional nanostructures in porous alumina. *Adv Funct Mater* 2008;18:2057-63
48. Zakeri R, Watts C, Wang HB, Kohli P. Synthesis and characterization of nonlinear nanopores in alumina films. *Chem Mater* 2007;19:1954-63
49. Losic D, Lillo M, Losic. Porous alumina with shaped pore geometries and complex pore architectures fabricated by cyclic anodization. *Small* 2009;5:1392-7
- **The first demonstration on controlling the shapes of AAO pores.**
50. Losic D. Losic DJr. Preparation of porous anodic alumina with periodically perforated pores. *Langmuir* 2009;25:5426-31
51. Popat KC, Mor G, Grimes CA, Desai TA. Surface modification of nanoporous alumina surfaces with poly(ethylene glycol). *Langmuir* 2004;20:8035-41
52. Losic D, Cole MA, Dollmann B, et al. Surface modification of nanoporous alumina membranes by plasma polymerization. *Nanotechnology* 2008;19: 245704(7 pg)
53. Cameron MA, Gartland IP, Smith JA, et al. Atomic layer deposition of SiO<sub>2</sub> and TiO<sub>2</sub> in alumina tubular membranes: Pore reduction and effect of surface species on gas transport. *Langmuir* 2000;16:7435-44
54. Bruening ML, Dotzauer DM, Jain P, et al. Creation of functional membranes using polyelectrolyte multilayers and polymer brushes. *Langmuir* 2008;24:7663-73
55. Velleman L, Triani G, Evans PJ, et al. Published online 12 June 2009. Microporous Mesoporous Mater 2009; DOI[SW2]:10.1016/j.micromeso.2009.05.024
56. Thormann A, Teuscher N, Pfannmoller M, et al. Nanoporous aluminum oxide membranes for filtration and biofunctionalization. *Small* 2007;3:1032-40
57. Chang CS, Suen SY. Modification of porous alumina membranes with n-alkanoic acids and their application in protein adsorption. *J Membr Sci* 2006;275:70-81
58. Bruening ML, Dotzauer DM, Jain P, et al. Creation of functional membranes using polyelectrolyte multilayers and polymer brushes. *Langmuir* 2008;24:7663-73
59. Darder M, Aranda P, Hernandez-Velez M, et al. Encapsulation of enzymes in alumina membranes of controlled pore size. *Thin Solid Films* 2006;495:321-6
60. Dai JH, Baker GL, Bruening ML. Use of porous membranes modified with polyelectrolyte multilayers as substrates for protein arrays with low nonspecific adsorption. *Anal Chem* 2006;78:135-40
61. Schmitt EK, Nurnabi M, Bushby RJ, Steinem C. Electrically insulating pore-suspending membranes on highly ordered porous alumina obtained from vesicle spreading. *Soft Matter* 2008;4:250-3

62. Jani AMM, Anglin EJ, McInnes SJP, et al. Nanoporous anodic aluminium oxide membranes with layered surface chemistry. *Chem Commun* 2009;21:3062-4
63. Kipke S, Schmid G. Nanoporous alumina membranes as diffusion controlling systems. *Adv Funct Mater* 2004;14:1184-8
64. Sedel L. Evolution of alumina-on-alumina implants. *Clinical orthopaedic and related research* 2000;379:48-54
65. Popat KC, Swan EEL, Mukhatyar V, et al. Influence of nanoporous alumina membranes on long-term osteoblast response. *Biomaterials* 2005;26:4516-22
- **Important study showing the interaction of porous alumina and osteoblast.**
66. Swan EEL, Popat KC, Grimes CA, Desai TA. Fabrication and evaluation of nanoporous alumina membranes for osteoblast culture. *J Biomed Mater Res. Part A* 2005;72A:288-95
67. La Flamme KE, Popat KC, Leoni L, et al. Biocompatibility of nanoporous alumina membranes for immunoisolation. *Biomaterials* 2007;28:2638-45
- **Important biocompatibility study of porous alumina for immunoisolation.**
68. Popat KC, Chatvanichkul KI, Barnes GL, et al. Osteogenic differentiation of marrow stromal cells cultured on nanoporous alumina surfaces. *J Biomed Mater Res Part A* 2007;80A:955-64
- **The first demonstration of osteogenic differentiation on porous alumina.**
69. Swan EEL, Popat KC, Desai TA. Peptide-immobilized nanoporous alumina membranes for enhanced osteoblast adhesion. *Biomaterials* 2005;26:1969-197
70. Karlsson M, Palsgard E, Wilshaw PR, Di Silvio L. Initial in vitro interaction of osteoblasts with nano-porous alumina. *Biomaterials* 2003;24:3039-46
71. Karlsson M, Johansson A, Tang L, Boman M. Nanoporous aluminum oxide affects neutrophil behaviour. *Microsc Research Technique* 2004;63:259-65
72. Karlsson M, Tang L. Surface morphology and adsorbed proteins affect phagocyte responses to nano-porous alumina. *J Mater Sci Mater Med* 2006;17:1101-11
73. Ferraz, N; Carlson J, Hong J, Ott MK. Influence of nanopore size on platelet adhesion and activation. *J Mater Sci Mater Med* 2008;19:3115-21
74. Jens D, Bose S, Hosick HL, Bandyopadhyay A. From CT scan to ceramic bone graft. *J Am Ceram Soc* 2003;86:1076-108
75. Karoussos IA, Wieneke H, Sawitowski T, et al. Inorganic materials as drug delivery systems in coronary artery stenting. *Materialwissenschaft und Werkstofftechnik* 2002;33:738-46
76. Kollum M, Farb A, Schreiber R, et al. Particle debris from a nanoporous stent coating obscures potential antiproliferative effects of tacrolimus-eluting stents in a porcine model of restenosis. *Catheterization and cardiovascular interventions* 2005;64:85-90
77. Sigler M; Paul T; Grabitz R G. Biocompatibility screening in cardiovascular implants. *Zeitschrift fuer Kardiologie* 2005;94:383-91
78. Camenzind E, DeScheerder IK. Local Drug delivery for coronary artery disease, Established and emerging applications. Oxford: Taylor & Francis 2005
79. Wieneke H, Sawitowski T, Wnendt S, et al. A new approach in interventional cardiology. *Herz* 2002;27:518-26
80. Wieneke H, Olaf Dirsch MD, Sawitowski T. Synergistic effects of a novel nanoporous stent coating and tacrolimus on intima proliferation in rabbits. *Catheterization and Cardiovascular Interventions* 2003;399-407
81. Tao SL, Desai TA. Microfabricated drug delivery systems. *Adv Drug Deliv Rev* 2003;55:315-28
- **Useful introductory review on micro and nanoporous devices for drug delivery.**
82. Desai TA. Bhatia S, Ferrari M. Therapeutic micro/nano technology. Berlin: Springer Sci 2006
- **The book describes the micro and nanoporous devices for drug delivery.**
83. Desai TA, West T, Cohen M, et al. Nanoporous microsystems for islet cell replacement. *Adv Drug Delivery Rev* 2004;56:1661-73
- **A review on nanoporous microsystems and devices.**
84. La Flamme KE, LaTempa TJ, Grimes CA, Desai TA. The effects of cell density and device arrangement on the behavior of macroencapsulated beta-cells. *Cell Transplantation* 2007;16:765-74
85. La Flamme KE, Gopal M, Gong D, Desai TA. Nanoporous Alumina Capsules for Cellular Macroencapsulation: Transport and Biocompatibility. *Diabetes Technol Therapeutics* 2005;7:684-694
- **A study demonstrating using AAO for immunoisolation.**
86. Gong D, Yadavalli V, V, et al. Therapeutic micro and nanotechnology: Controlled molecular release using nanoporous alumina capsules. *Biomedical Microdevices* 2003;5:75-80
- **A study demonstrating using AAO capsules for drug delivery.**
87. Zwilling V, Darque-Ceretti E, Boutry-Forveille A, et al. Structure and physicochemistry of anodic oxide films on titanium and TA6V alloy. *Surf Interfac Anal* 1999;27: 629-37
88. Kant K, Losic D. A simple approach for synthesis of TiO<sub>2</sub> nanotubes with through-hole morphology. *Phys Status Solidi RRL*, 2009;3:139-141
89. Macak JM, Albu SP, Schmuki P. Towards ideal hexagonal self-ordering of TiO<sub>2</sub> nanotubes. *Phys Status Solidi-RRL* 2007;1:181-3
90. Prakasam HE, Shankar K, Paulose M, et al. A new benchmark for TiO<sub>2</sub> nanotube array growth by anodization. *J Phys Chem C* 2007;111:7235-41
91. Paulose M, Peng L, Popat KC, et al. Fabrication of mechanically robust, large area, polycrystalline nanotubular/porous TiO<sub>2</sub> membranes. *J Membr Sci* 2008;319:199-205
92. Macak JM, Albu SP, Kim DH, et al. Multilayer TiO<sub>2</sub>-nanotube formation by two-step anodization. *Electrochem Solid State Lett* 2007;10:K28-31
93. Macak JM, Schmuki P. Anodic growth of self-organized anodic TiO<sub>2</sub> nanotubes in viscous electrolytes. *Electrochim Acta* 2006;52:1258-64
94. Bauer S, Kleber S, Schmuki P. TiO<sub>2</sub> nanotubes: Tailoring the geometry in H<sub>3</sub>PO<sub>4</sub>/HF electrolytes. *Electrochem Commun* 2006;8:1321
95. Albu SP, Kim D, Schmuki P. Growth of aligned TiO<sub>2</sub> bamboo-type nanotubes and highly ordered nanolace. *Angew Chem Int Ed* 2008;47:1916-9

96. Song YY, Schmidt-Stein F, Bauer S, Schmuki P. Amphiphilic TiO<sub>2</sub> Nanotube Arrays: An Actively Controllable Drug Delivery System. *J Am Chem Soc* 2009;131:4230-2
- **A demonstration of chemical modification of titania nanotubes and UV light triggering of drug release.**
97. Liu H, Webster TJ. Nanomedicine for implants. A review of studies and necessary experimental tools. *Biomaterials* 2007;28:354-69
98. Matsuno H, Yokoyama A, Watari F, et al. Biocompatibility and osteogenesis of refractory metal implants, titanium, hafnium, niobium, tantalum and rhenium. *Biomaterials* 2001;22:1253-1262
99. Kodama T. Study on biocompatibility of titanium alloys. *J Stomatol Soc Jpn* 1989;51:263-288
100. Popat KC, Leoni L, Grimes CA, Desai TA. Influence of engineered titania nanotubular surfaces on bone cells. *Biomaterials* 2007;28:3188-97
- **An important study showing the influence of topography on bone cell growth.**
101. Popat KC, Eltgroth M, LaTempa TJ, et al. Decreased Staphylococcus epidermis adhesion and increased osteoblast functionality on antibiotic-loaded titania nanotubes. *Biomaterials* 2009;28:4880-8
- **A study demonstrating drug delivery implant application of titania nanotubes.**
102. Popat KC, Eltgroth M, LaTempa TJ, et al. Titania nanotubes: A novel platform for drug-eluting coatings for medical implants. *Small* 2007;3:1878-81
103. Burns K, Yao C, Webster TJ. Increased chondrocyte adhesion on nanotubular anodized titanium. *J Biomed Mater Res. Part A* 2009;88A:561-8
104. Oh S, Daraio C, Chen LH, et al. Significantly accelerated osteoblast cell growth on aligned TiO<sub>2</sub> nanotubes. *J Biomed Mater Res. Part A* 2006;78A:97-103
- **The remarkable properties of titania nanotubes for osteoblast cell growth are demonstrated.**
105. Oh S, Jin S. Titanium oxide nanotubes with controlled morphology for enhanced bone growth. *Mater Sci Eng C* 2006; 26 C: 1301-1306
106. von Wilmsky C, Bauer S, Lutz R, et al. In Vivo Evaluation of Anodic TiO<sub>2</sub> Nanotubes: An Experimental Study in the Pig. *J Biomed Mater Res. Part B-Appl Biomater* 2009;89B:165-71
- **In vivo data demonstrating the potential of titania nanotube implants for bone growth.**
107. Oh S, Brammer KS, Li YSJ, et al. Stem cell fate dictated solely by altered nanotube dimension. *PNAS* 2009;106:2130-5
- **The demonstration of controllable osteogenic differentiation of hMSC stem cells by the pore size of titania nanotubes dimensions.**
108. Park J, Bauer S, von der Mark K, Schmuki P. Nanosize and vitality: TiO<sub>2</sub> nanotube diameter directs cell fate. *Nano Lett* 2007;7:1686-91
- **The first demonstration of controllable differentiation of stem cells by the pore size of titania nanotubes.**
109. Park J, Bauer S, Schlegel KA, et al. TiO<sub>2</sub> Nanotube Surfaces: 15 nm - An Optimal Length Scale of Surface Topography for Cell Adhesion and Differentiation. *Small* 2009;5:666-71
- **The demonstration of controllable cell differentiation and adhesion by the pore size of titania nanotubes.**
110. Bauer S, Park J, von der Mark K, Schmuki P. Improved attachment of mesenchymal stem cells on super-hydrophobic TiO<sub>2</sub> nanotubes. *Acta Biomaterialia* 2008;4:1576-82
- **A study showing the role of interfacial properties of titania nanotubes for the cell attachment.**
111. Peng LL, Mendelsohn AD, LaTempa TJ, et al. Long-Term small molecule and protein elution from TiO<sub>2</sub> nanotubes. *Nano Lett* 2009;9:1932-6
- **The first demonstration of application titania nanotubes for protein elution.**
112. Wykrzykowska JJ, Onuma Y, Serruys PW. Advances in stent drug delivery: the future is in bioabsorbable stents. *Expert Opin Drug Deliv* 2009;6:113-26
- **A review of stent drug delivery.**
113. Peng L, Eltgroth ML, LaTempa TJ, et al. The effect of TiO<sub>2</sub> nanotubes on endothelial function and smooth muscle proliferation. *Biomaterials* 2009;30:1268-72
114. Antoci V, Adams CS, Parvizi J, et al. The inhibition of Staphylococcus epidermidis biofilm formation by vancomycin-modified titanium alloy and implications for the treatment of periprosthetic infection. *Biomaterials* 2008;29:4684-90
115. Jose B, Antoci V, Zeiger AR, et al. Vancomycin covalently bonded to titanium beads kills Staphylococcus aureus. *Chem Biol* 2005;12:1041-8
- **The demonstration of drug attachment on the titania nanotube surface.**
116. Shrestha NK, Macak JM, Schmidt-Stein F, et al. Magnetically guided titania nanotubes for site-selective photocatalysis and drug release. *Angew Chem Int Ed* 2009;48:969-72
- **The first introduction of magnetically guided titania nanotubes and demonstration of their anticancer properties.**
117. Kalbacova M, Macak JM, Schmidt-Stein F, et al. TiO<sub>2</sub> nanotubes: photocatalyst for cancer cell killing. *Phys Status Solidi RRL* 2008;2:194-6
- **The first demonstration of anticancer properties of titania nanotube films.**
118. Farrar G, Altmann P, Watch S, et al. A. Defective gallium-transfer in binding in Alzheimer disease and Down syndrome: possible mechanism for accumulation of aluminium in brain. *Lancet* 1990;335:747-750
119. Goodman SB, Davidson JA, Fornasiero VL, et al. Histological response to cylinders of a low modulus titanium alloy (Ti-13Nb-13Zr) and a wear resistant zirconium alloy (Zr-2.5Nb) implanted in the rabbit tibia. *J Appl Biomater* 1993;4:331-339

## Affiliation

Dusan Losic<sup>†</sup> & Spomenka Simovic

<sup>†</sup>Author for correspondence

University of South Australia,

Ian Wark Research Institute,

Mawson Lakes Campus,

Mawson Lakes, Adelaide,

SA 5095, Australia

Tel: +61 8 8302 6862; Fax: +61 8 8302 3683;

E-mail: dusan.losic@unisa.edu.au